

Review Article

Molecular mechanism of α -tocopherol action

Angelo Azzi*

Vascular Biology Laboratory, Office 622, JM USDA-HNRCA at Tufts University, 711 Washington Street, Boston, MA 02111, USA

Received 19 October 2006; revised 1 March 2007; accepted 19 March 2007
Available online 24 March 2007

Abstract

The inability of other antioxidants to substitute for α -tocopherol in a number of cellular reactions, the lack of a compensatory antioxidant response in the gene expression under conditions of α -tocopherol deficiency, the unique uptake of α -tocopherol relative to the other tocopherols and its slower catabolism, and the striking differences in the molecular function of the different tocopherols and tocotrienols, observed in vitro, unrelated to their antioxidant properties, are all data in support of a nonantioxidant molecular function of α -tocopherol. Furthermore, in vivo studies have also shown that α -tocopherol is not able, at physiological concentrations, to protect against oxidant-induced damage or prevent disease allegedly caused by oxidative damage. α -Tocopherol appears to act as a ligand of not yet identified specific proteins (receptors, transcription factors) capable of regulating signal transduction and gene expression.

© 2007 Elsevier Inc. All rights reserved.

Contents

Definition	16
The chemical paradigm	16
The unverified extension of the antioxidant concept from chemistry into biology	17
Nonantioxidant physiological function of α -tocopherol: Evidence at a cellular level	17
Nonantioxidant physiological function of α -tocopherol: In vivo evidence	18
If α -tocopherol is not acting as an antioxidant what protects membrane phospholipid against oxidative damage?	18
A double role for α -tocopherol? Is DNA an antioxidant?	18
Conclusion.	19
References.	19

Definition

α -Tocopherol has been defined as a radical-chain breaker [1], which, due to its hydrophobic nature, operates in a lipid environment. The effects of α -tocopherol as an antioxidant are thus restricted to its direct effects in membranes and lipoprotein domains. Consequently, other definitions such as “secondary antioxidant,” antioxidant as inhibitor of “enzymes that produce radicals,” or activator of “genes coding for antioxidant enzymes” are confusing and do not help in understanding the

molecular mechanism of α -tocopherol function in vivo. The possible exclusion of α -tocopherol from the category of radical-chain breaker in a hydrophobic environment as defined above has prompted the reactive suggestion that the antioxidant properties of α -tocopherol may be exerted within a micro-domain of a receptor or of an enzyme. These types of suggestions, however, go beyond the discussion of the molecular aspects of α -tocopherol action.

The chemical paradigm

The antioxidant properties of α -tocopherol are a very well-established chemical paradigm. Indeed, vitamin E can act as an

* Fax: +1 617 556 3224.
E-mail address: angelo.azzi@tufts.edu.

antioxidant in the test tube, in lipid and phospholipid suspensions [1], in cell-free *Hevea brasiliensis* latex [2], or perhaps in plants, although in this case the alternative function of cellular signaling by modulating jasmonic acid levels has been also proposed [3]. There is little doubt that, *in vivo*, if given in pharmacological concentrations, possibly by parenteral administration to humans or animals, α -tocopherol must act as an antioxidant; however, this situation goes obviously beyond the concept of physiological function. Such an antioxidant function, that is, intrinsic part of the chemistry of the molecule, may in fact not be always desirable, similarly to the possible negative effects of the administration of other antioxidants in large amounts. It is known that the amount of the micronutrients, such as polyphenols, provided with an antioxidant function *in vitro*, which are accepted by the organism, is extremely low [4] and certainly below the amount needed to produce a significant antioxidant function. The emerging view is that polyphenols are likely to exert beneficial and/or toxic actions on cells not through their potential to act as antioxidants but rather through their modulation of protein and lipid kinase signaling cascades [5].

The unverified extension of the antioxidant concept from chemistry into biology

The argument that chemically tested antioxidants must have *in vivo* antioxidant properties is not tenable. Other *in vitro* “antioxidants” as ubiquinone [6] and carotenoids [7] have *in vivo* nonantioxidant properties. Also estrogens can be considered antioxidants [8], although not potent ones, and physiological levels of 17β -estradiol binding to LDL are associated with enhanced resistance to their Cu^{2+} -mediated oxidation [9]; however, this effect is not the consequence of radical scavenging; 17β -estradiol enhances the resistance of LDL to oxidation by stabilizing apoB-100 conformation [10]. In any case, 17β -estradiol, the most potent mammalian estrogenic hormone, is not acting by virtue of its antioxidant properties, but by binding to specific cellular receptors. Retinal [11], polyphenols, phytoestrogens, and flavonoids [4,12] are other examples of micronutrients provided with *in vitro* antioxidant capacity; the concentration that they reach *in vivo* is of the order of μM or lower that is not compatible with a significant *in vivo* antioxidant function [13]. Rather, by directly modulating signal transduction events they modify cell functional parameters [4]. An intriguing conjecture (there are not yet data to back it up) can be made at this point that the concentration of plant polyphenols provided with *in vitro* antioxidant properties is kept in the human organism extremely low by limiting their absorption and by induction of phase 1 and phase 2 enzymes, responsible for their modification, conjugation, and efficient elimination [13–15]. It appears in general as if diet antioxidant uptake must be avoided and that antioxidant concentration must be kept very low. The only exceptions appear to be ascorbic acid and α -tocopherol. It may not be surprising that natural selection has developed mechanisms intended to protect the organism from excessive antioxidant intake since reactive oxygen species have evolved as signaling molecules [16–21]. The activity of nox’s

(NADPH oxidases, present not only in macrophages but in a large number of nonphagocytosing cells) is tightly regulated by a number of enzymes [19,22,23] and is aimed at controlled production of superoxide and hydrogen peroxide. The latter is, for instance, capable of inhibiting protein tyrosine phosphatases with the consequent enhancement of the receptor tyrosine kinase signal [24–27]. Interference with such oxygen signaling by an antioxidant may not be desirable.

Nonantioxidant physiological function of α -tocopherol: Evidence at a cellular level

K.C.D Hickman wrote in 1946: “The cutting down of cell metabolism is a primary and intracellular function of vitamin E, and ... it has a secondary and more general antioxidant role which may be taken by other substances” [28] as cited in [29]. This conclusion was reached on the basis of the differential effects exhibited by vitamin E relative to methylene blue in preventing oxygen toxicity in the rat.

Such a conclusion was, in subsequent years, ignored with some exceptions such as A.T. Diplock who wrote in 1983 “The results suggest that α -tocopherol is capable of exerting a controlling influence upon the linoleyl and arachidonyl residues within membrane phospholipids which cannot be explained on the basis of the antioxidant function of the vitamin...” [30].

In more recent years the mechanism of action of α -tocopherol has been thoroughly reinvestigated. In light of new experimental findings, the view of Tappel [31] that the chain-breaking antioxidant vitamin E is the main protector against *in vivo* lipid peroxidation and of Burton and Ingold [32] that vitamin E functions in living systems primarily as a lipid antioxidant and free radical scavenger had to be revised. Among the important discoveries that have brought to this new paradigm is the finding that of the eight vitamin E family members (α -, β -, γ -, δ -tocopherol and the homonymous tocotrienols) only α -tocopherol (and to a much lesser extent γ -tocopherol) appears to be retained in significant amounts [33] by the organism. This event is the consequence of the expression in the liver of a protein, α -TTP, with high selectivity for α -tocopherol [34,35] and low or very low affinity for the other tocopherols with the implicit message of a particular evolutionary pressure exerted by α -tocopherol, which is not shared by other equally potent antioxidants. A second line of evidence comes from the experimental observations that α -tocopherol is able to modulate a number of cell functions in a unique way, not shared by any other antioxidants [36,37]. Our original observations were followed by a number of studies [38–44] indicating that a number of cell functions, such as inhibition of smooth muscle cell proliferation, preservation of endothelial integrity, inhibition of monocyte-endothelial adhesion, inhibition of monocyte reactive oxygen species and cytokine release, and inhibition of platelet adhesion and aggregation are controlled by the nonantioxidant properties of α -tocopherol. It is hard to imagine that such a fine regulation of cellular functions be mediated by noncontrollable free radical chain reactions [45]. After our original finding that α -tocopherol is able to modulate gene expression [46–50],

many other genes have been found to be under the control of α -tocopherol [51–56]. However, no genes expressing antioxidant enzymes appear to be up regulated in the absence of α -tocopherol as expected by an obvious compensatory mechanism. The different tocopherols and tocotrienols have effects, at a cellular level, that are independent of their relative antioxidant properties; for instance, the different tocopherols and the analogue compounds carbonitrile derivatives inhibit smooth muscle cell proliferation by a mechanism not correlated with the antioxidant properties of the molecules [57]. Moreover, the competition between α -tocopherol and β -tocopherol in inhibiting PKC or smooth muscle cell proliferation [58] suggests the existence of a common binding site for the two molecules and cannot be explained in terms of two antioxidants that, added together, have less effect than α -tocopherol alone. α -Tocotrienol has been also shown to act by the regulation of gene expression in an antioxidant-independent way [59]. γ -Tocopherol, less potent than α tocopherol as an antioxidant [60,61], has unique cellular functions, indicating again that their molecular structures and not their antioxidant properties determine the differential functions of tocopherols [62–72].

Nonantioxidant physiological function of α -tocopherol: In vivo evidence

In a number of in vivo situations, no antioxidant effect of α -tocopherol has been found. Only few of these observations, as examples, will be cited here. No effect of supplementation with vitamin E is seen on oxidative DNA damage as estimated by 8-oxo-7,8-dihydro-2'-deoxyguanosine excretion [73], again indicating that in vivo tocopherol did not act as an antioxidant. The fact that also vitamin C and coenzyme Q have no effect on 8-oxo-7,8-dihydro-2'-deoxyguanosine excretion [73] may raise a question regarding generalized antioxidant properties of these two molecules. Furthermore, antioxidants do not prevent muscle oxidative damage in response to an ultramarathon run [74]. Administration of vitamin E has been shown to trigger preconditioning via K(ATP) channels and cyclic-GMP without inhibiting lipid peroxidation [75]. Human atherosclerotic plaque contains both oxidized lipids and relatively large amounts of α -tocopherol and ascorbate, indicating that α -tocopherol is not able to act in the plaque, as an antioxidant [76]. Furthermore, human supplementation with α -tocopherol results in increased plasma and LDL tocopherol levels but the degree of protection against copper-catalyzed LDL oxidation is only evident at doses $>$ or $=$ 400 IU/day [73], thus indicating that, at physiological concentrations, no antioxidant effect can be demonstrated. α -Tocopherol does not have any protective effect against a number of pathologies, at the basis of which is presumably an excess of oxygen radical production such as on exercise-induced increases in muscle damage or recovery [74] or in carotid and aortic human lesions, where large amounts of oxidized lipids coexist with relatively normal α -tocopherol levels [76]. Also, recent mechanistic studies demonstrate that the role of α -tocopherol during the early stages of lipoprotein lipid peroxidation is complex and that the vitamin does not act as a chain-breaking antioxidant [77]. The poor performance of

antioxidant strategies using α -tocopherol in preventing either atherosclerosis or cardiovascular events is an established problem [78]. Such a situation casts severe doubts either on the implication of oxygen radicals as pathophysiological important for the onset of atherosclerosis [79] or on the in vivo efficacy of α -tocopherol as an antioxidant or on both [80].

If α -tocopherol is not acting as an antioxidant what protects membrane phospholipid against oxidative damage?

A number of compounds produced physiologically in the body in a much regulated way have been shown to act in protecting membranes against lipid oxidation. Among them, bilirubin has been shown to be an antioxidant of physiological importance [81] whose production is regulated by the oxidant-inducible enzyme heme oxygenase.

Superoxide radicals can also reduce membrane damage by acting as radical chain breakers [82] as well as nitric oxide, which has been shown to react with lipid peroxy radicals exhibiting great oxidant protection [83]. Finally, phospholipid-hydroperoxide glutathione peroxidase (GPx-4) is a well-established mechanism for phospholipid hydroperoxide repair [84].

A double role for α -tocopherol? Is DNA an antioxidant?

The fact that α -tocopherol plasma or tissue concentration may be diminished under conditions of high radical production (sepsis, smoking, etc) and its oxidation products may be excreted has been taken as evidence that α -tocopherol acts as an antioxidant. However, excretion of oxidized α -tocopherol products does not imply that α -tocopherol has finalistically sacrificed itself to protect the organism against free radicals. In fact, similarly to DNA, α -tocopherol requires protection by other antioxidant systems to prevent its loss and with it, the loss of its regulatory properties. When the oxidative mechanisms are not compensated by sufficient protective mechanisms, burning up of α -tocopherol may take place with the appearance of its oxidation products; parallelly, changes in the signaling effects of the molecule may take place. It is well known that an excess of free radicals can produce DNA single-strand and double-strand breaks and the appearance in the urine of base oxidation products such as 8-OH guanine. It is also known that this damage can be repaired, with great efficiency, by appropriate mechanisms. If the specific function of DNA and its hierarchical superiority relative to all other cell functions were not known, DNA could be considered a mechanism for free radical scavenging, equipped with a capable recycling mechanism.

It appears that the relationship between α -tocopherol and its oxidative environment is that of a sensor, monitoring the environment and, through its concentration changes, transferring information to the cell. Recycling phenomena and TAP (tocopherol-associated proteins) protection [85], as a consequence of the tight interaction between protein and α -tocopherol [86], may be mechanisms of preserving α -tocopherol from oxidative damage and degradation. Given the

specific functions of α -tocopherol, it is unthinkable to attribute to α -tocopherol a co-primary role as antioxidant: if α -tocopherol were an antioxidant, its concentration would diminish as a consequence of an increased radical production with abrogation of important physiological functions. As discussed above, α -tocopherol must be protected against free radical damage rather than be used to eliminate free radicals. “Recycling” is therefore intended to regenerate damaged α -tocopherol and not to reactivate a lost antioxidant.

Conclusion

A number of lines of evidence, evolutionary, genetic, biochemical, and functional, have indicated that the natural function of α -tocopherol is that of cell signaling. Such a property is not shared by any other antioxidant molecule. Recent experiments have indicated that α -tocopherol, but not other antioxidants, is the precursor of a more active form of vitamin E, α -tocopheryl phosphate; this species may be ultimately the molecule which specifically interacts with a receptor or transcription factor and modulates cell functions [87,88]. α -Tocopherol has been shown as well not to protect in vivo against oxidative damage or to prevent diseases which have at their basis an oxidative insult. Altogether, the conclusion can be drawn that α -tocopherol is not physiologically acting as an antioxidant.

References

- [1] Barclay, L. R. C.; Ingold, K. U. Autoxidation of biological molecules: 2. The autoxidation of a model membrane. A comparison of the autoxidation of egg lecithin phosphatidylcholine in water and in chlorobenzene. *J. Am. Chem. Soc.* **103**:6478–6485; 1981.
- [2] Whittle, K. J.; Audley, B. G.; Pennock, J. F. The incorporation of [14C] methionine into chromanols and quinones by *Hevea brasiliensis* latex. *Biochem. J.* **103**:21C–22C; 1967.
- [3] Munne-Bosch, S.; Weiler, E. W.; Alegre, L.; Muller, M.; DUCHTING, P.; Falk, J. Alpha-tocopherol may influence cellular signaling by modulating jasmonic acid levels in plants. *Planta* **225**:681–691; 2006.
- [4] Manach, C.; Williamson, G.; Morand, C.; Scalbert, A.; Remesy, C. Bioavailability and bioefficacy of polyphenols in humans: I. Review of 97 bioavailability studies. *Am. J. Clin. Nutr.* **81**:230S–242S; 2005.
- [5] Williams, R. J.; Spencer, J. P.; Rice-Evans, C. Flavonoids: antioxidants or signalling molecules? *Free Radic. Biol. Med.* **36**:838–849; 2004.
- [6] Crane, F. L.; Navas, P. The diversity of coenzyme Q function. *Mol. Aspects Med.* **18**:S1–S6; 1997.
- [7] Stahl, W.; Ale-Agha, N.; Polidori, M. C. Non-antioxidant properties of carotenoids. *Biol. Chem.* **383**:553; 2002.
- [8] Ayres, S.; Tang, M.; Subbiah, M. T. Estradiol-17 β as an antioxidant: some distinct features when compared with common fat-soluble antioxidants. *J. Lab. Clin. Med.* **128**:367–375; 1996.
- [9] Shwaery, G. T.; Vita, J. A.; Keaney, J. F. Jr. Antioxidant protection of LDL by physiological concentrations of 17 beta-estradiol. Requirement for estradiol modification. *Circulation* **95**:1378–1385; 1997.
- [10] Brunelli, R.; Mei, G.; Krasnowska, E. K.; Pierucci, F.; Zichella, L.; Ursini, F. Parasassi T. Estradiol enhances the resistance of LDL to oxidation by stabilizing apoB-100 conformation. *Biochemistry* **39**:13897–13903; 2000.
- [11] Keys, S. A.; Zimmerman, W. F. Antioxidant activity of retinol, glutathione, and taurine in bovine photoreceptor cell membranes. *Exp. Eye Res.* **68**:693–702; 1999.
- [12] Manson, M. M.; Ball, H. W.; Barrett, M. C.; Clark, H. L.; Judah, D. J.; Williamson, G.; Neal, G. E. Mechanism of action of dietary chemoprotective agents in rat liver: induction of phase I and II drug metabolizing enzymes and aflatoxin B1 metabolism. *Carcinogenesis* **18**:1729–1738; 1997.
- [13] Scalbert, A.; Williamson, G. Dietary intake and bioavailability of polyphenols. *J. Nutr.* **130**:2073S–2085S; 2000.
- [14] Williamson, G.; Manach, C. Bioavailability and bioefficacy of polyphenols in humans: II. Review of 93 intervention studies. *Am. J. Clin. Nutr.* **81**:243S–255S; 2005.
- [15] Kroon, P. A.; Clifford, M. N.; Crozier, A.; Day, A. J.; Donovan, J. L.; Manach, C.; Williamson, G. How should we assess the effects of exposure to dietary polyphenols in vitro? *Am. J. Clin. Nutr.* **80**:15–21; 2004.
- [16] Li, J.; Stouffs, M.; Serrander, L.; Banfi, B.; Bettiol, E.; Charnay, Y.; Steger, K.; Krause, K. H.; Jaconi, M. E. The NADPH oxidase NOX4 drives cardiac differentiation: role in regulating cardiac transcription factors and MAP kinase activation. *Mol. Biol. Cell* **17**:3978–3988; 2006.
- [17] Schmelter, M.; Ateghang, B.; Helmig, S.; Wartenberg, M.; Sauer, H. Embryonic stem cells utilize reactive oxygen species as transducers of mechanical strain-induced cardiovascular differentiation. *FASEB J.* **20**:1182–1184; 2006.
- [18] Bell, R. M.; Cave, A. C.; Johar, S.; Hearse, D. J.; Shah, A. M.; Shattock, M. J. Pivotal role of NOX-2-containing NADPH oxidase in early ischemic preconditioning. *FASEB J.* **19**:2037–2039; 2005.
- [19] Wolin, M. S.; Ahmad, M.; Gupte, S. A. Oxidant and redox signaling in vascular oxygen sensing mechanisms: basic concepts, current controversies, and potential importance of cytosolic NADPH. *Am. J. Physiol. Lung Cell Mol. Physiol.* **289**:L159–L173; 2005.
- [20] Lambeth, J. D. NOX enzymes and the biology of reactive oxygen. *Nat. Rev. Immunol.* **4**:181–189; 2004.
- [21] Jackson, S. H.; Devadas, S.; Kwon, J.; Pinto, L. A.; Williams, M. S. T cells express a phagocyte-type NADPH oxidase that is activated after T cell receptor stimulation. *Nat. Immunol.* **5**:818–827; 2004.
- [22] Finkel, T.; Holbrook, N. J. Oxidants, oxidative stress and the biology of ageing. *Nature* **408**:239–247; 2000.
- [23] Sen, C. K. Redox signaling and the emerging therapeutic potential of thiol antioxidants. *Biochem. Pharmacol.* **55**:1747–1758; 1998.
- [24] Meng, T. C.; Buckley, D. A.; Galic, S.; Tiganis, T.; Tonks, N. K. Regulation of insulin signaling through reversible oxidation of the protein-tyrosine phosphatases TC45 and PTP1B. *J. Biol. Chem.* **279**:37716–37725; 2004.
- [25] Meng, T. C.; Tonks, N. K. Analysis of the regulation of protein tyrosine phosphatases in vivo by reversible oxidation. *Methods Enzymol.* **366**:304–318; 2003.
- [26] Salmeen, A.; Andersen, J. N.; Myers, M. P.; Meng, T. C.; Hinks, J. A.; Tonks, N. K.; Barford, D. Redox regulation of protein tyrosine phosphatase 1B involves a sulphenyl-amide intermediate. *Nature* **423**:769–773; 2003.
- [27] Meng, T. C.; Fukada, T.; Tonks, N. K. Reversible oxidation and inactivation of protein tyrosine phosphatases in vivo. *Mol. Cell* **9**:387–399; 2002.
- [28] Hickman, K. C. D. Tocopherol interrelationship. *Adv. Enzymol.* **6**:469–524; 1946.
- [29] Taylor, D. W. The effects of vitamin E and of methylene blue on the manifestations of oxygen poisoning in the rat. *J. Physiol.* **27**:200–206; 1956.
- [30] Diplock, A. T. The role of vitamin E in biological membranes. *Ciba Found. Symp.* **101**:45–55; 1983.
- [31] Tappel, A. L. Vitamin E and selenium protection from in vivo lipid peroxidation. *Ann. N. Y. Acad. Sci.* **355**:18–31; 1980.
- [32] Burton, G. W.; Joyce, A.; Ingold, K. U. First proof that vitamin E is major lipid-soluble, chain-breaking antioxidant in human blood plasma [letter]. *Lancet* **2**:327; 1982.
- [33] Traber, M. G.; Kayden, H. J. Preferential incorporation of alpha-tocopherol vs gamma-tocopherol in human lipoproteins. *Am. J. Clin. Nutr.* **49**:517–526; 1989.
- [34] Sato, Y.; Hagiwara, K.; Arai, H.; Inoue, K. Purification and characterization of the alpha-tocopherol transfer protein from rat liver. *FEBS Lett.* **288**:41–45; 1991.

- [35] Hosomi, A.; Arita, M.; Sato, Y.; Kiyose, C.; Ueda, T.; Igarashi, O.; Arai, H.; Inoue, K. Affinity for alpha-tocopherol transfer protein as a determinant of the biological activities of vitamin E analogs. *FEBS Lett.* **409**:105–108; 1997.
- [36] Mahoney, C. W.; Azzi, A. Vitamin E inhibits protein kinase C activity. *Biochem. Biophys. Res Commun.* **154**:694–697; 1988.
- [37] Boscoboinik, D.; Szewczyk, A.; Hensey, C.; Azzi, A. Inhibition of cell proliferation by alpha-tocopherol. Role of protein kinase C. *J. Biol. Chem.* **266**:6188–6194; 1991.
- [38] Cecchini, T.; Ciaroni, S.; Ferri, P.; Ambrogini, P.; Cuppini, R.; Santi, S.; Del Grande, P. Alpha-tocopherol, an exogenous factor of adult hippocampal neurogenesis regulation. *J. Neurosci. Res.* **73**:447–455; 2003.
- [39] Hacquebard, M.; Carpentier, Y. A. Vitamin E: absorption, plasma transport and cell uptake. *Curr. Opin. Clin. Nutr. Metab. Care* **8**:133; 2005.
- [40] Miyoshi, N.; Naniwa, K.; Kumagai, T.; Uchida, K.; Osawa, T.; Nakamura, Y. Alpha-tocopherol-mediated caspase-3 up-regulation enhances susceptibility to apoptotic stimuli. *Biochem. Biophys. Res. Commun.* **334**:466–473; 2005.
- [41] Rimbach, G.; Minihane, A. M.; Majewicz, J.; Fischer, A.; Pallauf, J.; Virgli, F.; Weinberg, P. D. Regulation of cell signalling by vitamin E. *Proc. Nutr. Soc.* **61**:415–425; 2002.
- [42] Ruch, R. J.; Bandyopadhyay, S.; Somani, P.; Klaunig, J. E. Evaluation of amiodarone free radical toxicity in rat hepatocytes. *Toxicol. Lett.* **56**:117–126; 1991.
- [43] van Aalst, J. A.; Burmeister, W.; Fox, P. L.; Graham, L. M. Alpha-tocopherol preserves endothelial cell migration in the presence of cell-oxidized low-density lipoprotein by inhibiting changes in cell membrane fluidity. *J. Vasc. Surg.* **39**:229–237; 2004.
- [44] Devaraj, S.; Jialal, I. The effects of alpha-tocopherol on critical cells in atherosclerosis. *Curr. Opin. Lipidol.* **9**:11–15; 1998.
- [45] Yamamoto, Y.; Niki, E.; Eguchi, J.; Kamiya, Y.; Shimasaki, H. Oxidation of biological membranes and its inhibition. Free radical chain oxidation of erythrocyte ghost membranes by oxygen. *Biochim. Biophys. Acta* **819**:29–36; 1985.
- [46] Ricciarelli, R.; Azzi, A. Regulation of recombinant PKC alpha activity by protein phosphatase 1 and protein phosphatase 2A. *Arch. Biochem. Biophys.* **355**:197–200; 1998.
- [47] Ricciarelli, R.; Zingg, J. M.; Azzi, A. Vitamin E reduces the uptake of oxidized LDL by inhibiting CD36 scavenger receptor expression in cultured aortic smooth muscle cells. *Circulation* **102**:82–87; 2000.
- [48] Villacorta, L.; Graca-Souza, A. V.; Ricciarelli, R.; Zingg, J. M.; Azzi, A. Alpha-tocopherol induces expression of connective tissue growth factor and antagonizes tumor necrosis factor-alpha-mediated downregulation in human smooth muscle cells. *Circ. Res.* **92**:104–110; 2003.
- [49] Ricciarelli, R.; Maroni, P.; Ozer, N.; Zingg, J. M.; Azzi, A. Age-dependent increase of collagenase expression can be reduced by alpha-tocopherol via protein kinase C inhibition. *Free Radic. Biol. Med.* **27**:729–737; 1999.
- [50] Azzi, A.; Gysin, R.; Kempna, P.; Munteanu, A.; Villacorta, L.; Visarius, T.; Zingg, J. M. Regulation of gene expression by alpha-tocopherol. *Biol. Chem.* **385**:585–591; 2004.
- [51] Barella, L.; Muller, P. Y.; Schlachter, M.; Hunziker, W.; Stocklin, E.; Spitzer, V.; Meier, N.; de Pascual-Teresa, S.; Minihane, A. M.; Rimbach, G. Identification of hepatic molecular mechanisms of action of alpha-tocopherol using global gene expression profile analysis in rats. *Biochim. Biophys. Acta* **1689**:66–74; 2004.
- [52] Rota, C.; Barella, L.; Minihane, A. M.; Stocklin, E.; Rimbach, G. Dietary alpha-tocopherol affects differential gene expression in rat testes. *IUBMB Life* **56**:277–280; 2004.
- [53] Rimbach, G.; Fischer, A.; Stoecklin, E.; Barella, L. Modulation of hepatic gene expression by alpha-tocopherol in cultured cells and in vivo. *Ann. N. Y. Acad. Sci.* **1031**:102–108; 2004.
- [54] Hyland, S.; Muller, D.; Hayton, S.; Stoecklin, E.; Barella, L. Cortical gene expression in the vitamin E-deficient rat: possible mechanisms for the electrophysiological abnormalities of visual and neural function. *Ann. Nutr. Metab.* **50**:433–441; 2006.
- [55] Gohil, K.; Godzdzank, R.; O'Roark, E.; Schock, B. C.; Kaini, R. R.; Packer, L.; Cross, C. E.; Traber, M. G. Alpha-tocopherol transfer protein deficiency in mice causes multi-organ deregulation of gene networks and behavioral deficits with age. *Ann. N. Y. Acad. Sci.* **1031**:109–126; 2004.
- [56] Gohil, K.; Schock, B. C.; Chakraborty, A. A.; Terasawa, Y.; Raber, J.; Farese, R. V. Jr.; Packer, L.; Cross, C. E.; Traber, M. G. Gene expression profile of oxidant stress and neurodegeneration in transgenic mice deficient in alpha-tocopherol transfer protein. *Free Radic. Biol. Med.* **35**:1343–1354; 2003.
- [57] Boscoboinik, D.; Özer, N. K.; Moser, U.; Azzi, A. Tocopherols and 6-hydroxy-chroman-2-carbonitrile derivatives inhibit vascular smooth muscle cell proliferation by a nonantioxidant mechanism. *Arch. Biochem. Biophys.* **318**:241–246; 1995.
- [58] Tasinato, A.; Boscoboinik, D.; Bartoli, G. M.; Maroni, P.; Azzi, A. d-Alpha-tocopherol inhibition of vascular smooth muscle cell proliferation occurs at physiological concentrations, correlates with protein kinase C inhibition, and is independent of its antioxidant properties. *Proc. Natl. Acad. Sci. USA* **92**:12190–12194; 1995.
- [59] Roy, S.; Lado, B. H.; Khanna, S.; Sen, C. K. Vitamin E sensitive genes in the developing rat fetal brain: a high-density oligonucleotide microarray analysis. *FEBS Lett.* **530**:17–23; 2002.
- [60] Pryor, A. W.; Cornicelli, J. A.; Devall, L. J.; Tait, B.; Trivedi, B. K.; Witak, D. T.; Wu, M. A rapid screening test to determine the antioxidant potencies of natural and synthetic antioxidants. *J. Org. Chem.* **58**:3521–3532; 1993.
- [61] Xu, Z.; Hua, N.; Godber, J. S. Antioxidant activity of tocopherols, tocotrienols, and gamma-oryzanol components from rice bran against cholesterol oxidation accelerated by 2,2'-azobis(2-methylpropanamide) dihydrochloride. *J. Agric. Food Chem.* **49**:2077–2081; 2001.
- [62] Itoh, N.; Masuo, Y.; Yoshida, Y.; Cynshi, O.; Jishage, K.; Niki, E. Gamma-tocopherol attenuates MPTP-induced dopamine loss more efficiently than alpha-tocopherol in mouse brain. *Neurosci. Lett.* **403**:136–140; 2006.
- [63] Takahashi, K.; Komaru, T.; Takeda, S.; Takeda, M.; Koshida, R.; Nakayama, M.; Kokusho, Y.; Kawakami, Y.; Yamaguchi, N.; Miyazawa, T., et al. Gamma-tocopherol, but not alpha-tocopherol, potently inhibits neointimal formation induced by vascular injury in insulin resistant rats. *J. Mol. Cell. Cardiol.* **41**:544–554; 2006.
- [64] Samandari, E.; Visarius, T.; Zingg, J. M.; Azzi, A. The effect of gamma-tocopherol on proliferation, integrin expression, adhesion, and migration of human glioma cells. *Biochem. Biophys. Res. Commun.* **342**:1329–1333; 2006.
- [65] Campbell, S. E.; Stone, W. L.; Lee, S.; Whaley, S.; Yang, H.; Qui, M.; Goforth, P.; Sherman, D.; McHaffie, D.; Krishnan, K. Comparative effects of RRR-alpha- and RRR-gamma-tocopherol on proliferation and apoptosis in human colon cancer cell lines. *BMC Cancer* **6**:13; 2006.
- [66] Hensley, K.; Benaksas, E. J.; Bolli, R.; Comp, P.; Grammas, P.; Hamdheydari, L.; Mou, S.; Pye, Q. N.; Stoddard, M. F.; Wallis, G., et al. New perspectives on vitamin E: gamma-tocopherol and carboxyethyl-hydroxychroman metabolites in biology and medicine. *Free Radic. Biol. Med.* **36**:1–15; 2004.
- [67] Campbell, S. E.; Stone, W. L.; Whaley, S. G.; Qui, M.; Krishnan, K. Gamma (gamma) tocopherol upregulates peroxisome proliferator activated receptor (PPAR) gamma (gamma) expression in SW 480 human colon cancer cell lines. *BMC Cancer* **3**:25; 2003.
- [68] Jiang, Q.; Ames, B. N. Gamma-tocopherol, but not alpha-tocopherol, decreases proinflammatory eicosanoids and inflammation damage in rats. *FASEB J.* **17**:816–822; 2003.
- [69] Devaraj, S.; Traber, M. G. Gamma-tocopherol, the new vitamin E? *Am. J. Clin. Nutr.* **77**:530–531; 2003.
- [70] Mishima, K.; Tanaka, T.; Pu, F.; Egashira, N.; Iwasaki, K.; Hidaka, R.; Matsunaga, K.; Takata, J.; Karube, Y.; Fujiwara, M. Vitamin E isoforms alpha-tocotrienol and gamma-tocopherol prevent cerebral infarction in mice. *Neurosci. Lett.* **337**:56–60; 2003.
- [71] Gysin, R.; Azzi, A.; Visarius, T. Gamma-tocopherol inhibits human cancer cell cycle progression and cell proliferation by down-regulation of cyclins. *FASEB J.* **16**:1952–1954; 2002.
- [72] Zhang, P.; Omaye, S. T. Antioxidant and prooxidant roles for beta-carotene, alpha-tocopherol and ascorbic acid in human lung cells. *Toxicol. In Vitro* **15**:13–24; 2001.
- [73] Prieme, H.; Loft, S.; Nyyssonen, K.; Salonen, J. T.; Poulsen, H. E. No effect of supplementation with vitamin E, ascorbic acid, or coenzyme Q10

- on oxidative DNA damage estimated by 8-oxo-7,8-dihydro-2'-deoxyguanosine excretion in smokers. *Am. J. Clin. Nutr.* **65**:503–507; 1997.
- [74] Mastaloudis, A.; Traber, M. G.; Carstensen, K.; Widrick, J. J. Antioxidants did not prevent muscle damage in response to an ultramarathon run. *Med. Sci. Sports Exerc.* **38**:72–80; 2006.
- [75] Andreadou, I.; Iliodromitis, E. K.; Tsovolas, K.; Aggeli, I. K.; Zoga, A.; Gaitanaki, C.; Paraskevaïdis, I. A.; Markantonis, S. L.; Beis, I.; Kremastinos, D. T. Acute administration of vitamin E triggers preconditioning via K(ATP) channels and cyclic-GMP without inhibiting lipid peroxidation. *Free Radic. Biol. Med.* **41**:1092–1099; 2006.
- [76] Suarna, C.; Dean, R. T.; May, J.; Stocker, R. Human atherosclerotic plaque contains both oxidized lipids and relatively large amounts of alpha-tocopherol and ascorbate. *Arterioscler. Thromb. Vasc. Biol.* **15**:1616–1624; 1995.
- [77] Stocker, R.; Keane, J. F. Jr. New insights on oxidative stress in the artery wall. *J. Thromb. Haemost.* **3**:1825–1834; 2005.
- [78] Ueda, S.; Yasunari, K. What we learnt from randomized clinical trials and cohort studies of antioxidant vitamin? focus on vitamin E and cardiovascular disease. *Curr. Pharm. Biotechnol.* **7**:69–72; 2006.
- [79] Stocker, R.; Keane, J. F. Jr. Role of oxidative modifications in atherosclerosis. *Physiol. Rev.* **84**:1381–1478; 2004.
- [80] Halliwell, B. Phagocyte-derived reactive species: salvation or suicide? *Trends Biochem. Sci.* **31**:509–515; 2006.
- [81] Stocker, R.; McDonagh, A. F.; Glazer, A. N.; Ames, B. N. Antioxidant activities of bile pigments: biliverdin and bilirubin. *Methods Enzymol.* **186**:301–309; 1990.
- [82] Kowald, A.; Lehrach, H.; Klipp, E. Alternative pathways as mechanism for the negative effects associated with overexpression of superoxide dismutase. *J. Theor. Biol.* **238**:828–840; 2006.
- [83] Rubbo, H.; Radi, R.; Anselmi, D.; Kirk, M.; Barnes, S.; Butler, J.; Eiserich, J.; Freeman, B. Nitric oxide reaction with lipid peroxy radicals spares alpha-tocopherol during lipid peroxidation. Greater oxidant protection from the pair nitric oxide/alpha-tocopherol than alpha-tocopherol/ascorbate. *J. Biol. Chem.* **275**:10812–10818; 2000.
- [84] Januel, C.; El Hentati, F. Z.; Carreras, M.; Arthur, J. R.; Calzada, C.; Lagarde, M.; Vericel, E. Phospholipid-hydroperoxide glutathione peroxidase (GPx-4) localization in resting platelets, and compartmental change during platelet activation. *Biochim. Biophys. Acta* **1761**:1228–1234; 2006.
- [85] Zimmer, S.; Stocker, A.; Sarbolouki, M. N.; Spycher, S. E.; Sassoon, J.; Azzi, A. A novel human tocopherol-associated protein: cloning, in vitro expression, and characterization. *J. Biol. Chem.* **275**:25672–25680; 2000.
- [86] Yamauchi, J.; Iwamoto, T.; Kida, S.; Masushige, S.; Yamada, K.; Esashi, T. Tocopherol-associated protein is a ligand-dependent transcriptional activator. *Biochem. Biophys. Res. Commun.* **285**:295–299; 2001.
- [87] Negis, Y.; Zingg, J. -M.; Ogru, E.; Gianello, R.; Libinaki, R.; Azzi, A. On the existence of cellular tocopheryl phosphate, its synthesis, degradation and cellular roles: a hypothesis. *IUBMB Life* **57**:23–25; 2005.
- [88] Negis, Y.; Aytan, N.; Ozer, N.; Ogru, E.; Libinaki, R.; Gianello, R.; Azzi, A.; Zingg, J. M. The effect of tocopheryl phosphates on atherosclerosis progression in rabbits fed with a high cholesterol diet. *Arch. Biochem. Biophys.* **450**:63–66; 2006.